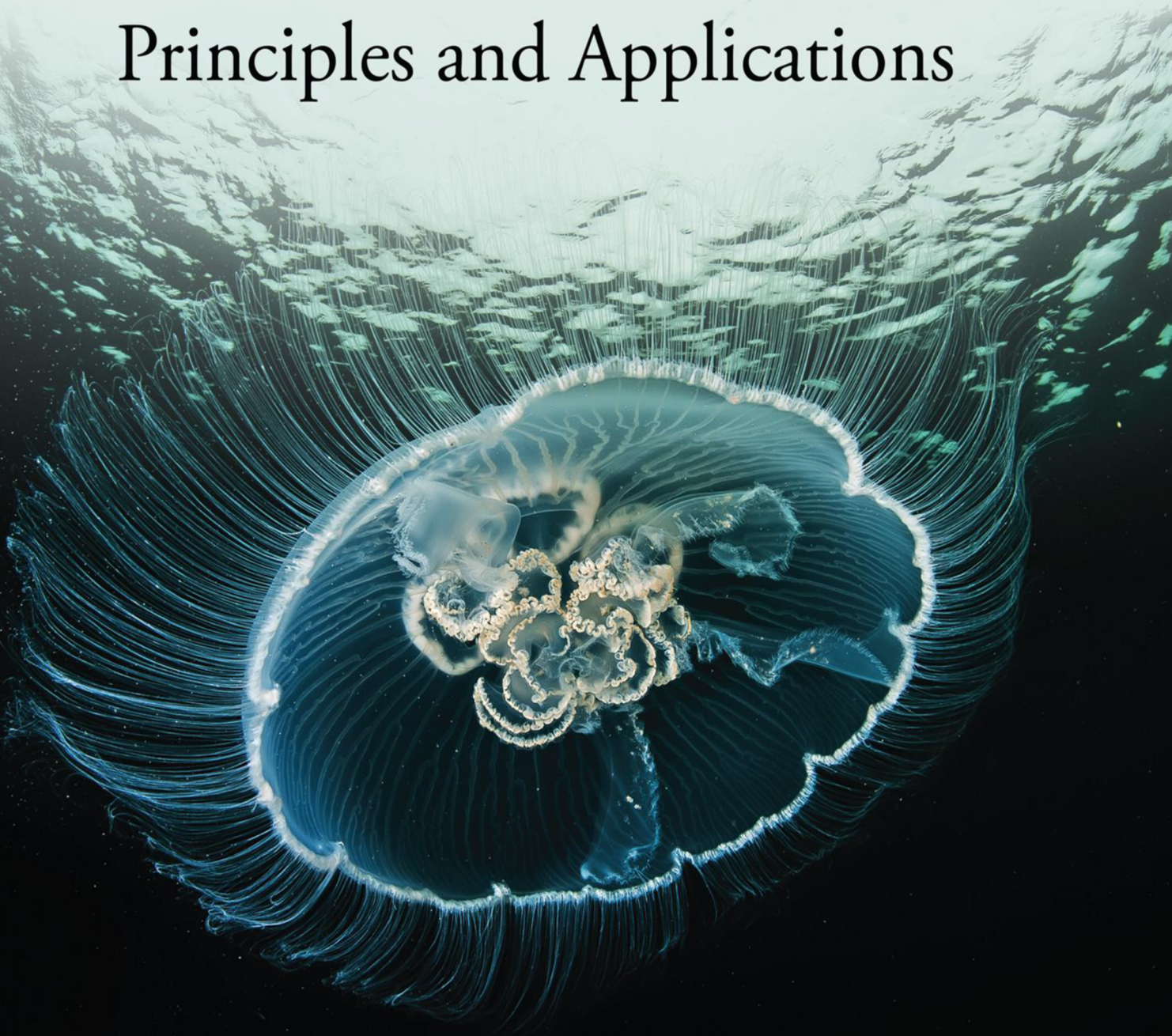


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MARINE GLYCOBIOLOGY

Principles and Applications



edited by
Se-Kwon Kim

**MARINE
GLYCOBIOLOGY**
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Preface

Marine glycobiology is the study of carbohydrate and carbohydrate with molecules (protein, lipids, enzymes, or small molecules). Glycoconjugates are glycan linked with other biological molecules. The study and research on marine glycobiology is less well-known. However, the recent development on analytical instruments and chemical characterizations increases the research on glycoconjugates. The knowledge gained of the exact chemical structure of marine glycoconjugates increases its use in biological and biomedical applications.

This book contains 38 chapters under different sections.

1. Section I—Chapter 1 provides a general introduction to the topics covered in this book.
2. Section II—marine glycoconjugates of reproduction and chemical communications (Chapters 3 and 4) are described.
3. Section III—Bioactivity, principles, and applications of marine glycans (Chapters 5 through 7) are explored.
4. Section IV—marine glycoproteins (Chapters 8 through 13)—deals with biomedical benefits of algal glycoproteins, marine source collagen, and detoxification in the marine environment.
5. Section V—marine glycoenzymes (Chapter 14)—discusses the sialyltransferases from the marine environment and their applications.
6. Section VI—marine carbohydrates (Chapters 15 through 29)—discusses several marine carbohydrates and their glycobiology. Marine bacterial exopolysaccharides, agar, chitin and chitosan, sulfated polysaccharides, carbohydrates from marine microbes, algal polysaccharides, and mangroves are discussed in detail. In addition, the use of these polysaccharides in pharmaceutical applications, plant growth, and various biological activities are also discussed.
7. Section VII and VIII—bioinformatics and biological role of glycoconjugates (Chapters 30 and 31)—describes glycan's predictive modeling using modern algorithms and glycoconjugated bioactivity compounds and biological applications.
8. Section IX—glycoconjugates as biomedicine (Chapters 32 through 38)—presents the application of glycoconjugates in biomedicine and their biotechnological applications.

I express my sincere thanks to all the authors who have contributed toward this book. Their relentless effort was the result of their strong inclination towards scientific research, and great perseverance descended from their experience. I am grateful to the experts who have contributed to this book.

I hope that fundamental as well as applied contributions to this book might serve as potential research and development leads for the benefit of humankind. Marine glycobiology will be the excellent field in the future towards the enrichment of targeted marine glycans, which further sets up a suitable for further applications. This book would be a reference book for students in academic and industrial research.

Prof. Se-Kwon Kim
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this book for marine biotechnology and glycobiology researchers/students/industrialists and hope that it helps to enhance their understanding in this field.

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Se-Kwon Kim, PhD, is a distinguished professor in the Department of Marine Bio Convergence Science and Technology and as director of Marine Bioprocess Research Center (MBPRC) at Pukyong National University, Busan, South Korea.

He received his MSc and PhD from Pukyong National University and conducted his postdoctoral studies in the Laboratory of Biochemical Engineering, University of Illinois, Urbana–Champaign, Illinois. Later, he became a visiting scientist at the Memorial University of Newfoundland and the University of British Columbia in Canada.

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His major research interests are investigation and development of bioactive substances from marine resources. His immense experience in marine bioprocessing and mass production technologies for marine bio-industry is the key asset of holding majorly funded marine bio projects in Korea. Furthermore, he expended his research fields up to the development of bioactive materials from marine organisms for their applications in oriental medicine, cosmeceuticals, and nutraceuticals. To date, he has authored around 700 research papers and 70 books and holds 130 patents.



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Agar-abundant marine carbohydrate from seaweeds in Indonesia

Production, bioactivity, and utilization

Syamdidi, Hari Eko Irianto and Giyatmi Irianto

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18.1 Introduction

Agar is one of the marine carbohydrates and, as a polysaccharide complex, obtained through bleaching and hot water extraction of agarocytes from the red alga Rhodophyceae. There are two genera of seaweeds used as agar sources for agar industries in Indonesia: *Gelidium* and *Gracilaria*. In general, *Gelidium*, *Acanthopeltis*, *Ceramium*, *Pterocladia*, and *Gracilaria* are predominant raw materials in agar production in the world. Agar consists of about 70% agarose and 30% agaropectin (Scott and Eagleson 1998). Agarose is a neutral gelling fraction, which consists of a linear polymer of alternating D-galactose and 3,6-anhydrogalactose units. Agaropectin is a non-gelling fraction, which consists of 1,3 glycosidically linked D-galactose units, some of which are sulfated at position 6. Chemical structures of agarose and agaropectin are shown in [Figure 18.1](#).

Seaweed is abundant in Indonesia, of which around 555 species have been already identified. About 21 species have been utilized as raw material by seaweed

processing industries (Aslan 1991). Tremendous development of seaweed farming during the past 5 years has brought about the situation in which the seaweed production of Indonesia (in volume) is higher than in the Philippines; the production volume in 2013 reached 9,298,474 tons (Ministry of Marine Affairs and Fisheries 2014). Most of them are mainly *Eucheuma* sp. and *Gracilaria* sp.

Red algae are the most popular seaweeds that are used for food, pharmaceutical, and other industries. From 17 genera of red algae, 34 species have already been used for various purposes. Moreover, 23 species are able to be cultured: 6 species from the genus *Eucheuma*, 3 species from *Gelidium*, 10 species from *Gracilaria*, and 4 species from *Hypnea*. The genus *Eucheuma*, *Gracilaria*, and *Gelidium* are commonly found and cultured in Indonesian waters. However, only the genera producing agar (agarophytes) and carrageenan (carrageenophytes) are commercially cultured to support the seaweed industry in Indonesia and also to fulfill worldwide demand (Kordi 2011).

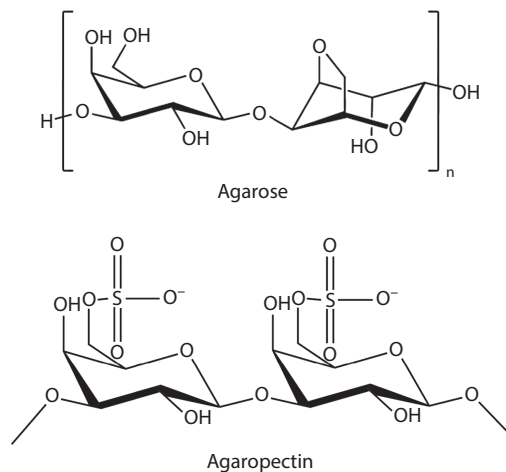


Figure 18.1 Structure of agarose (1,4)-3,6 anhydro L-galactose, (1,3) D-galactose, and agarpectin. (From Ramadhan, W., Utilization of agar powder as texturizer in guava (*Psidium guajava* L.) spread sheet and shelf-life prediction, Faculty of Fisheries and Marine Science, Bogor Agricultural University, Bogor, Indonesia, 2011. With permission.)

Many countries have developed methods of agar processing, but only two methods are widely used: the freeze-thaw method, and the pressing method. Indonesia is currently the largest agar producer in the world. Agar produced by Indonesian processing factories is not only used for domestic consumption but also export to several countries, for example, China, Japan, and European countries.

Several investigations have been conducted by researchers all over the world to reveal the health benefits of agar, the findings of which are expected to guide into new prospective utilization. Some research has shown that the bioprospecting substances of agar have the potential to alleviate health problems. Therefore, new value-added products based on those findings are expected to be developed in the future, and Indonesia is poised to play an important role due to the abundance resources of agar-producing seaweeds as raw material.

18.2 Agar industries in Indonesia

World trade of agar in terms of raw material of agar and finished products shows an increasing trend. The world demand was estimated to be 10,000 tons/year of raw material and 3,500 tons/year of finished products (Directorate General of Aquaculture 2005). Indonesia can develop the agar industry in three directions: as a raw material supplier, an agar producer, or both. Agar production looks more strategic for Indonesia, because the industry can contribute to the welfare of people by providing them job opportunities.

Agar industry is growing rapidly in Indonesia. Production of agar in Indonesia started before the World War II. In 1930, the first agar factory was established in Kudus, Central Java. This was followed by others in Jakarta, Surabaya, and Makassar (Sulistyo 2002). PT Sinar Kencana, an agar factory in Surabaya, East Java, was founded in 1947. In 1955, there were five companies operating with a total production around 13.7 tons per annum (tpa). After four decades, 10 new agar companies were established with the total production reaching 108.7 tpa. Thus the number of agar producing companies became 15 in 1993 (Zatnika 1997) but currently only 11 companies are existing. One of them is the largest agar company in the world with total production of over 3000 tpa.

18.2.1 Agar source

Red algae genera such as *Gelidium*, *Gracilaria*, *Grateloupia*, *Halymenia*, *Hypnea*, and *Porphyra* (Table 18.1) are used as raw material in agar production in Indonesia. They are naturally available throughout the Indonesian waters. Currently, the demand of *Gracilaria* has grown to supply raw material for agar processing because of the increasing demand of agar and for establishing new agar factories. Seaweed from the wild cannot fulfill raw material need of the agar industry, which has led to the exploration of another source of raw material, farmed seaweeds. Culturing *Gracilaria* was developed later on around the year 1980 to ensure continuous supply of raw material for the agar industry. *Gracilaria* is now widely cultured by the farmers along the coast, particularly in Sulawesi and Java. One of the biggest *Gracilaria* culture ponds is located in Tangerang, Banten Province.

18.2.2 Seaweed harvesting

Seaweed is normally harvested after 45 days if cultured in the beach but after 60–75 days if cultured in ponds (Kordi 2011). The Directorate General of Aquaculture (2005) suggested that *Gracilaria* is harvested after 90 days of cultivation, and the next harvesting can be performed after 60 days. Harvesting should be performed at the right age to obtain the optimum yield of agar during processing. Harvesting age also affects the quality of the extracted agar, in which underage seaweed will result in a lower agar quality.

Harvested *Gracilaria* is then washed with fresh water to free it from unwanted materials including other seaweed species. Seaweeds are subsequently sun-dried by placing them on drying racks, mats, or floors. The drying step should not be delayed so as to avoid deterioration due to fermentation. Drying can take 3–4 days till achieving a moisture content of about 25% as required by the market. While waiting for marketing, dried

Table 18.1 Seaweeds used as agar source in Indonesia

Seaweed genera	Location in Indonesia
<i>Gelidium amansii</i>	Alor Islands, Tanimbar Islands, Maluku Islands
<i>G. rigidium</i>	Scattered across Indonesian coast
<i>G. latifolium</i>	Bengkulu, Lampung, South part of Java Island, West Nusa Tenggara Islands
<i>Gracilaria confervoides</i>	Scattered across Indonesian coast
<i>G. verrucosa</i>	West Sumbawa, Sawu Island, South Sulawesi, Southeast Sulawesi
<i>G. euchemoides</i>	South Lampung, South Java Island, Southeast Celebes, South and Southeast Moluccas
<i>G. lichenoides</i>	Scattered across Indonesian coast
<i>G. gigas</i>	Scattered across Indonesian coast
<i>G. taenoides</i>	Riau Islands, Bangka and Belitung Islands, Lampung
<i>Grateloupia filicina</i>	West and South Java Island, South Lampung, Seribu Islands
<i>Halymenia durvillei</i>	South and Southeast Celebes, Ambon and Seram Islands, Papua, East Nusa Tenggara, Lombok, Sumbawa and Halmahera
<i>Hypnea cervicornis</i>	Riau Islands, Bali, Tawi-tawi
<i>H. divacirata</i>	Riau Islands, Moluccas Islands
<i>H. musciformis</i>	Scattered across Indonesian coast
<i>Porphyra atropurpurea</i>	Halmahera Island and Kei Island

Source: Anggadiredja, J.T. et al., *Seaweed: Culturing, Processing and Marketing Potential Fisheries Commodity*, Penebar Swadaya, Jakarta, Indonesia, 2006.

seaweed is stored for a certain period. According to Rodarte et al. (2010), the agar content of *Gracilaria* from the tropics decreases in a few months because of hydrolysis. The hydrolysis in *Gracilaria* could be due to the presence of agarolytic bacteria, of which the most important is *Bacillus cereus*, and to the presence of the algae's own agarolytic enzyme. Storage of *Gracilaria cornea* for 2 years and *Gracilaria euchematoides* for 1 year resulted in a reduction of a gel strength of 17% and 35%, respectively.

18.2.3 Agar extraction

In Indonesia, there are three types of agar products in the market: agar sheet, agar bar, and agar powder. Basically, the extraction methods used to make those products are similar.

Agar is extracted from *Gracilaria* by means of two steps of cooking using water, in which the ratio of dried seaweed and water is approximately 1:20. The first cooking is carried out with the dried seaweed and an agar/water ratio of 1:14 at 85°C–95°C and pH 6–7 for 2 h. The agar extract and seaweed pulp are separated using fabrics. The seaweed pulp obtained is boiled again for the second cooking with the dried seaweed and with a water ratio of 1:6 for 1 h. The agar extract is then added with 2%–3% KOH or KCl for gel formation (Directorate General of Aquaculture 2005). The gel undergoes further processing for producing agar sheet, agar bar, or agar powder by applying specific treatments for each product.

Mostly, farmed *Gracilaria* produces a soft-textured gel resulting in difficulties for further processing,

particularly for industries operated on a small scale. To improve the gel properties, the farmed *Gracilaria* is mixed with that harvested from the nature before agar extraction (Aji et al. 2003).

Some authors (Rao and Bekheet 1976; Chapman and Chapman 1980; Robello et al. 1997; Montano et al. 1999) have found that soaking the seaweed in an alkali solution can improve the gel strength of agar. The chemical structures of the agar precursor in seaweed and of the idealized agar after alkali treatment are shown in Figure 18.2. Kusuma et al. (2013) have shown that the concentration of NaOH affected the gel strength, sulfate content, ash content, moisture content, and yield

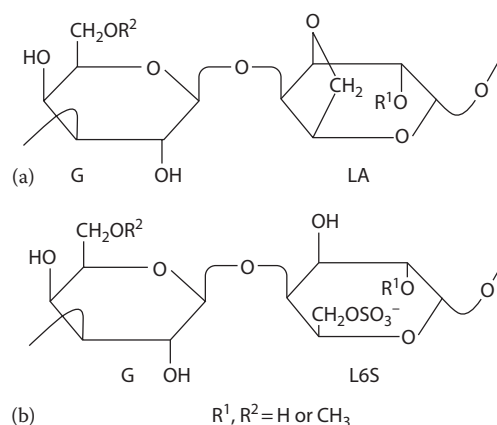


Figure 18.2 Chemical structure of agar. (a) Idealized agar. (b) Agar precursor prior to extraction (From Distantina, S., et al., *Jurnal Rekayasa Proses*, 2, 11, 2008. With permission.)

of agar *G. verrucosa*. The higher the concentration of NaOH, the higher the gel strength, the ash content, the water content, and the yield, but the lower the sulfate levels. The best agar quality was obtained using 6% NaOH solution. Distantina et al. (2008) and Van et al. (2008) noted that soaking seaweeds in an alkali solution resulted in lower extraction rate and lower yield, but higher gel strength compared to soaking in an acid solution.

18.2.4 Agar quality

The quality of Indonesian agar is regulated under National Standardization Board. This institution regulates the specification of agar for both safety and wholesomeness (Table 18.2). Most agar industries in Indonesia have no problem meeting the standards required by the Board. Moreover, they usually sell the products above requirement standard, especially for moisture content, which ranges from 10% to 15%. International standards

Table 18.2 Indonesian agar specification

Parameter	Unit	Requirement
<i>Sensory</i>	—	Min. 7 (1–9 scale) ^a
<i>Chemical quality</i>		
Moisture content	%	Max. 22
Ash content ^b	%	Max. 6.5
Acid-insoluble ash ^b	%	Max. 0.5
Starch ^b	—	Negative
Gelatin and protein ^b	—	Negative
<i>Microbial quality</i>		
TPC	Colony/g	Max. 5000
<i>Escherichia coli</i>	APM/g	<3
Salmonella	Per 25 g	Negative
Yeast and moulds	Colony/g	Max. 300
<i>Heavy metals^b</i>		
Arsenic (As)	mg/kg	Max. 3
Cadmium (Cd)	mg/kg	Max. 1
Mercury (Hg)	mg/kg	Max. 1
Lead (Pb)	mg/kg	Max. 3
Zinc (Zn)	mg/kg	Max. 40
<i>Physic^b</i>		
Water absorption	—	Min. five times
Foreign insoluble matter	%	Max. 1
Particle size (pass 60 mesh size)	%	Min. 80

Source: National Standardization Board, *SNI Agar Powder*, BSN, Jakarta, Indonesia, 2014.

^a For each sensory attribute.

^b If only requested.

published by, for example, by the EU (European Union) and JECFA (The Joint FAO/WHO Expert Committee on Food Additives) are quite similar to the Indonesian standard. However, in industrial trading, moisture content, color, and odor are the most demanding parameters. Other parameters are conditional, depending on the buyer's requirements.

18.3 Bioactivity of agar

Studies have been carried out to reveal the bioactivity of agar-producing seaweeds, not specifically agar itself. Some species of red algae were reported to exhibit a broad spectrum of biological activities. The bioactivity of the most common agar seaweeds in Indonesia—from genus *Gracilaria* and *Gelidium*—has been examined. Almeida et al. (2011) have excellently reviewed the bioactivity of *Gracilaria* from some studies that highlight the potential pharmaceutical uses, particularly in terms of toxic, cytotoxic, spermicidal, anti-implantation, antibacterial, antiviral, antifungal, antiprotozoa, antihypertensive, antioxidant, anti-inflammatory, analgesic, and spasmolytic effects in the gastrointestinal tract.

18.3.1 Anti-inflammatory

Anti-inflammatory activities have been reported for a sulfated polysaccharide fraction from *Gracilaria caudate* (Chavez et al. 2013), a galactan from *Gelidium crinale* (Sousa et al. 2013), and polysaccharide fractions from *G. verrucosa* (Yoshizawa et al. 1996). Sajiki (1997) reported that *G. verrucosa*, *G. asiatica*, *G. lichenoides* and other species contain PGE₂, which produce physiological effects including hyperthermia, hypotension, smooth-muscle dilatation, hyperalgesia, and gastric secretion inhibition (Minghetti and Levi 1998).

18.3.2 Contraception activity

A methanol/methylene chloride (1:1) extract from *G. corticata* exhibited post-coital contraceptive activity due to enhanced pre-implantation without any marked side effects when introduced orally to female rats. This shows that red marine algae are a potential source for post-coital contraceptive drugs. Extracts from *G. edulis* demonstrated 100% inhibition of sperm motility, and this effect was related to the disruption of the plasma membrane by spermicidal compounds (Almeida et al. 2011).

18.3.3 Antioxidant

The antioxidant activity of *Gracilaria* is related to the anti-inflammatory effects. Swantara and Parwata (2011) found that *G. coronopifolia* has 90.27% antioxidant activity consisting of 1-nonadecane, hexadecanoic acid,

9-octadecanoic acid, cholest-4,6-dien-3-ol, cholest-5-en-3 β -ol (cholesterol), and cholest-4-en-3-one. Murugan and Iyer (2012) noted that *G. edulis* possesses antioxidant activity.

18.3.4 Gastrointestinal and cardiovascular effects

Aqueous extract from dried *G. verrucosa* or fresh *G. chorda* at a dose of 0.5 mg per animal was able to control gastrointestinal disorders in mice, the active compounds being zeaxanthin, antheraxanthin, carotenoids, pyrimidine 2-amino-4-carboxy, non-alkaloid nitrogen heterocycle, steroids, 5- α -poriferastane, 3- β -6- α -diol poriferastane, 5- α -3- β -6- β -diol, and gigatinine. Meanwhile, a 90% ethanol extract from *G. edulis* showed diuretic activity, and an aqueous extract from *G. lichenoides* administered intravenously showed antihypertensive effect in rats (Almeida et al. 2011).

18.3.5 Antibiotic activity

Gracilaria corticata were found to be active against gram-positive cultures of *Bacillus* (Bhakuni and Rawat 2005). The authors also found that an extract of *Gelidium cartilagineum* and *Chondrus crispus* were active against influenza B and mumps virus. This activity has been attributed to the presence of a galactan unit in the polysaccharides, agar, and carrageenan present in both species.

Another genus of *Gracilaria*—*Gracilaria* Greville—contains a gelatinous nontoxic colloidal carbohydrate in the cell wall and intercellular spaces of the algae and has wide use in the preparation of food, ice creams, jellies, soups, bacteriological samples, and cosmetics (Gosh et al. 2012; Kerton et al. 2013). They are sources of important bioactive metabolites with antibiotic activity (Smit 2004).

A chloroform extract of *G. edulis* (Gmelin) Silva was found to have antibacterial activity against some bacterial strains, in which the isolated steroids (carotenoids, β -cryptoxanthin, and β -carotene) and carbohydrates are suspected as the active compounds. An ethanol extract from *G. debilis* showed antibacterial activity against *Staphylococcus aureus*. A 95% ethanol extract from whole dried *G. cervicornisalgae* was active against *S. aureus* at a concentration of 5.0 mg/mL. A methanol extract from fresh *G. corticata* was active against *Bacillus subtilis*, *B. megaterium*, *S. aureus*, and *Streptococcus viridans*. Ethanol extracts from *G. domingensis* and *G. sjoestedii* showed antibacterial activity against *E. coli* and *S. aureus*. Ethanol extracts from *G. debilis*, *G. domingensis*, and *G. sjoestedii* were active against *Candida albicans*. An ethanol extract from *G. domingensis* was active against *Mycobacterium smegmatis* and *Neurospora crassa*. *G. domingensis* has as chemical

constituents polysaccharide CT-1, palmitic acid, and steroids (stigmasterol, sitosterol, campesterol, cholest-7-en-3- β -ol, and brassicasterol) (Almeida et al. 2011).

18.3.6 Antiviral activity

Extracts from *G. bursa-pastoris* and *Gracilaria* sp. were inactive against the herpes simplex 1 virus (HSV) and the human immunodeficiency virus (HIV) when evaluated in cell cultures. Granin BP and citrullinyl-arginine proteins were isolated from these extracts. A methanol extract from dried *G. pacifica* at a concentration of 200.0 μ g/mL was active against Sindbis virus, but was not effective against HSV when tested at a concentration of 400 μ g/mL. Extracts and compounds obtained from *Gracilaria* sp. with anti-HIV activity are also active against other retroviruses such as HSV (Almeida et al. 2011).

18.4 Agar utilization

The many uses of agar and agarose are related to the formation of thermoreversible gels at low concentration in water with large hysteresis. The physicochemical and rheological properties of these algal polysaccharides are linked to their chemical structure (Lahaye and Rochas 1991). Agar has been in use as a source of food in some Asian countries for a long time. Most of agar utilization is for food (~80%–90%) and the rest is used for biotechnological applications (FAO 2003; Armisen and Galatas 2009). Based on their types, agar was divided into natural agar and industrial agar. Natural agar was produced by traditional method while the industrial agar using freezing-defreezing techniques by artificial freezing, or accelerated syneresis through pressure (Armisen 1995) (Table 18.3).

Agar is used as a thickening agent and also to improve the texture of processed food such as jellies, dairy products, fruit pastilles, chewing gum, canned meats, soups, confectionery, and baked goods and icings, as well as frozen and salted fish (Armisen and Galatas 2009). Its neutral taste and high fiber content make agar superior to other hydrocolloid additives. These properties have led to broadening the application of agar to many food products. Agar is generally safe for human consumption. According to the U.S. Food and Drug Administration (FDA), agar is classified as GRAS (Generally Recognized as Safe), as an E406 additive by the European Commission, and is registered as 9002-18-0 in the Register Service of the Chemical Abstracts (Armisen and Galatas 2009).

In future, agar is expected to be widely used in biotechnology application because of its benefits. About 10% of the production is classified as too low for a GRAS product. Agar consists of agarose, which is a

Table 18.3 Agar grades, their application and plant origins

	Agar type applications	Type of seaweed
Natural agar	“Strip” “Square”: accustomed only in Far East traditional kitchen	Produced mostly with <i>Gelidium</i> by traditional methods
Industrial agar	Food-grade agar used for industrial food production Pharmacological agar Clonic plants production grade Bacteriological grade used for bacteriological culture media formulation Purified agar used in biochemistry and in culture media for very difficult bacteria	<i>Gelidium</i> , <i>Gracilaria</i> , <i>Gracilariopsis</i> , <i>Pterocladia</i> , <i>Ahnfeltia</i> , <i>Gelidiella</i> <i>Gelidium</i> <i>Gelidium</i> or <i>Pterocladia</i> <i>Gelidium</i> or <i>Pterocladia</i> <i>Gelidium</i>

Source: Armisen, R., *J. Appl. Phycol.*, 7, 231, 1995. With permission.

neutral polysaccharide, and agarpectin, which is polysaccharide sulfate. As a gelling agent, agar is widely used in the food, pharmaceutical, and cosmetics industries. Agarose is widely used in the field of biotechnology, both as a culture medium and an electrophoresis medium. Pure agarose is one of main ingredients of microbial and plant culture media (Istini et al. 2001).

The possible use of agar in food application is related to its properties: its gel-forming capability and its unique, reversible gelling performance. In the food industry, agar mostly used as gelling agent for bread, jelly, candies, dairy products, and ice cream. The food industry has been growing fast during the last decades in line with the increasing world population. Food trend nowadays is changing to healthy products that are low in fat but high in fiber. Agar contains high fiber, which can be used in food fortification to increase health benefit due to its indigestibility by the human metabolic system. Compared to carrageenan, agar is composed of soluble fibers since its content is above 94%, which is higher than that of carrageenan (67%–80%) (Armisen and Galatas 2009). Murata and Nakazoe (2001) found that agar consumption led to a decrease in the concentration of blood glucose and caused an anti-aggregation effect on red blood cells. In the beverage industry, agar is used in beer, coffee, and wine purifying process and also as an emulsifier for chocolate products. Agar with a concentration 0.1%–1% can be used as an emulsifier for some products such as yoghurt, cheese, jelly, and bakery products.

Agar consists of two fractions: agarose and agarpectin (Armisen and Galatas 1987). Agarose has low sulfate content and hence mostly used for biotechnology application (Wahyuni 2003). Agarose is used as an ingredient of making gels for electrophoresis of protein

isolation and purification. In addition, agarose is used in chromatography columns, which has been commercialized production with brand Sepharose (Pharmacia) and Bio-Gel A (Bio-Rad). Agarose also has been widely used in up to 250,000 Da for particle separation as well as for virus, protein, and chromosome separation (Wahyuni 2003).

18.5 Conclusions

Indonesia has abundance resources for marine carbohydrate production, especially red algae as a source of agar. However, the existing utilization of those resources is mainly for the production of conventional food products, namely agar powder. Recent exploration has revealed other benefits of agar, which is actually not only to fulfill human consumption needs but also to overcome some health problems based on the bioactive substances contained in agar-producing seaweeds. A new use for agar by taking advantage of its bioactive property is to generate products with significantly higher added value instead of producing conventional products. It is expected that the high added value can be realized by all stakeholders involved in the agar industry, including seaweed farmers whose role is often neglected. This way, the goals of the Indonesian government of improving the national welfare and alleviating poverty through seaweed development can be achieved as planned.

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